Total synthesis of myxothiazols, novel bis-thiazole β -methoxyacrylate-based anti-fungal compounds from myxobacteria[†]

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Received 7th March 2006, Accepted 9th June 2006 First published as an Advance Article on the web 29th June 2006 DOI: 10.1039/b603433k

Convergent total syntheses of myxothiazols A and Z are described. The syntheses are based on elaboration of the (*S*)-*E*,*E*-diene thioamide **22**, conversion of **22** into the bis-thiazole **27** and Wittig reactions between **27c** and the aldehyde **30**. The substituted β -methoxyacrylate aldehyde **30** was produced *via* an Evans asymmetric aldol protocol or *via* the 2H-pyran-2-one **31**. An *E*-selective Wittig reaction between the ylide derived from the phosphonium salt **27c** and the (+)-aldehyde **30** led to (+)-myxothiazol Z (**1b**), and a corresponding reaction with the (±)-acrylamide aldehyde **44** gave (±)-myxothiazol A (**1a**). Complementary studies led to synthesis of the ester **47b**, corresponding to myxothiazol R and myxothiazol S.

Introduction

Myxothiazol is the generic name used to describe a family of fungicides isolated from myxobacteria, *e.g. Myxococcus fulvus*, *Angiococcus disciformis*, and characterised by the presence of a novel and unusual β -methoxyacrylate pharmacophore linked to a 2,4-disubstituted bis-thiazole unit. Myxothiazol A (**1a**) was the first member to be isolated, in 1980,¹ and myxothiazol Z (**1b**) was described in the primary literature in 1999.² In 1988, Hőfle and Sakagami and their respective co-workers³ reported a different group of β -methoxyacrylate fungicides from various species of myxobacteria which were named melithiazols, *e.g.* melithiazol A (**2**), and cystothiazols, *e.g.* **3a** and **3b**. Interestingly, the secondary metabolites **1**, **2**, and **3** are related to the strobilurins, *e.g.* **4**, and the oudemansins, *e.g.* **5**, produced by various fungi.⁴

All of the aforementioned β -methoxyacrylates act as inhibitors of mitochondrial respiration by blocking electron transfer between cytochrome b and cytochrome c₁.⁵ An exciting range of synthetic analogues of strobilurin has now been brought to the market as agricultural fungicides,⁶ and synthetic studies within the melithiazols and cystothiazols have been vigorous in recent years.⁷ Before these recent endeavours, however, in 1993, we described the first, and only, synthesis of (±)-myxothiazol A (1a).⁸ Since this time, we have carried out further synthetic investigations towards the myxothiazols, and in this paper we draw together these studies, culminating in total syntheses of several of their members.

Results and discussion

Synthetic strategies

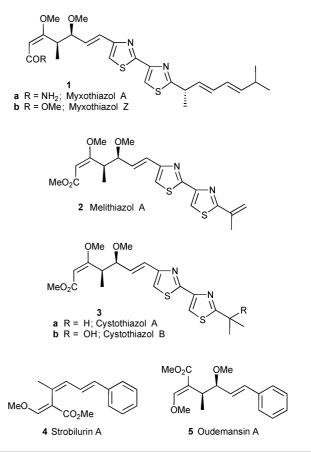
The myxothiazol structure **1** accommodates a substituted β methoxyacrylate left-hand side, *viz.* **6**, linked to a bis-thiazole by an *E*-butenyl unit, carrying two stereogenic centres. The bis-thiazole

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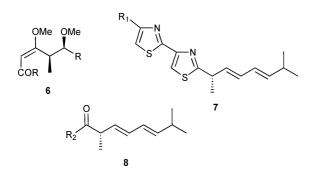
† Electronic supplementary information (ESI) available: experimental details. See DOI: 10.1039/b603433k

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in 1 is further substituted on the right-hand side by a conjugated E,E-heptadienyl unit, *viz.* 8, carrying a third stereogenic centre. The presence of these structural units in the myxothiazols 1 suggested a straightforward strategy to their synthesis based on (i) synthesis of the E,E-heptadienyl side chain, 8, followed by (ii) conversion into the substituted bis-thiazole 7, and finally (iii) linking the left-hand side β -methoxyacrylate residue 6. In our



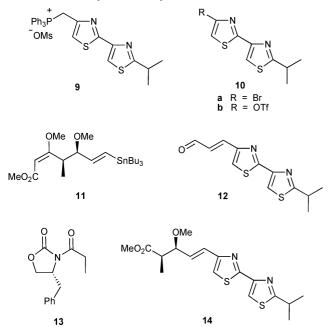
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earlier studies,⁸ we synthesised the bis-thiazole unit 7 and attached the left-hand side chain in the structure 1 using a Wittig reaction between the ylide derived from the bis-thiazole phosphonium salt 7 and the aldehyde 6 (R = CHO). This strategy was later

$(R_1 = CH_2 \stackrel{+}{P} Ph_3 I)$

applied by Charette and Deroy^{7b} and by Akita and co-workers^{7a} in their syntheses of cystothiazols A (**3a**) and B (**3b**) using the phosphonium salt **9**. In other studies Bach and Heuser^{7d} developed a Suzuki cross-coupling reaction from the 4-bromo bis-thiazole **10a** to synthesise cystothiazol E, and Shao and Panek,^{7e} in 2004, applied the Stille cross-coupling reaction between **10b** and **11** in their synthesis of cystothiazols. By contrast, Williams *et al.*^{7c} produced the left-hand side chain in their synthesis of cystothiazols A and C using an asymmetric aldol reaction between the α , β unsaturated aldehyde **12** and the Evans oxazolidinone **13**, leading ultimately to **14**. This general strategy was also later used by Ojika *et al.*^{7f} in another synthesis of cystothiazol A.



In our own studies, we explored a number of complementary pathways to elaborate the interesting substituted β methoxyacrylate side chain, *viz.* **6**, in the myxothiazols. These methods, alongside procedures to synthesise the right-hand side chain **8** attached to the bis-thiazole unit **7**, together with the assembly of these units leading to syntheses of myxothiazol A (**1a**) and Z (**1b**), and also the methyl ester corresponding to myxothiazols R and S, will now be described.

The (S)-E,E-heptadienyl side chain 8

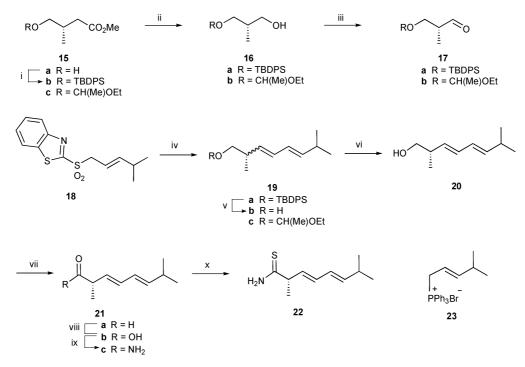
The heptadienyl side chain in the myxothiazols, appropriately functionalised as a thioamide, i.e. 22, was conveniently synthesised starting from commercially available (R) methyl 3-hydroxy-2methylpropionate 15a (Scheme 1). Thus, protection of 15a as its TBDPS ether, followed by reduction of the ester group in 15b, to the corresponding carbinol 16a and oxidation first gave the aldehyde 17a as a low-melting solid. A Julia olefination reaction⁹ between the aldehyde 17a and the E-benzothiazole sulfone 18 derived from E-4-methylpent-2-en-1-ol,¹⁰ in the presence of sodium bis(trimethylsilylamide) at -78 °C next led to the conjugated diene 19a, which was produced as a 4 : 1 mixture of E- and Zisomers at the newly introduced alkene bond. Deprotection of the TBDPS group in 19a, followed by treatment of the resulting diene alcohol 19b (4: 1 mixture of E-3, E-5 and Z-3, E-5 isomers) with iodine in refluxing diethyl ether under ultraviolet light irradiation, and chromatography, gave the geometrically pure E-3, E-5 diene alcohol 20 as a colourless liquid. A Mosher's ester analysis gave an ee > 98%,¹¹ and the E-3, E-5 geometry of the conjugated diene followed conclusively from examination of the magnitude of the vicinal couplings for the olefinic protons in the ¹H NMR spectrum of 20. The same diene alcohol 20 was also obtained via a Wittig reaction between the l-ethoxyethyl derivative 17b, and the phosphonium salt 23, using *n*-BuLi in diethyl ether at -10 °C, which first gave the conjugated diene 19c, also as a 4 : 1 mixture of E-3, E-5 and Z-3, E-5 isomers. Deprotection of 19c, followed by iodine-catalysed equilibration of the resulting diene alcohol then gave the E-3, E-5 diene alcohol 20 in a similar overall yield as the alternative Julia olefination protocol. The E-3, E-5 diene alcohol 20 was next smoothly converted into the corresponding thioamide 22 in four straightforward steps, *i.e.* oxidation to the carboxylic acid 21b using Dess-Martin periodinane followed by buffered sodium chlorite; conversion of the acid 21b into the amide 21c and, finally, treatment of 21c with Lawesson's reagent in THF.

The substituted bis-thiazole 7

A useful variety of methods is now available for making 2,4disubstituted thiazoles, including the Hantzsch method¹² described as early as 1887, and a more biomimetic approach which proceeds *via* thiazoline intermediates produced from cysteinyl peptide precursors.¹³ Both of these general methods can be used in an iterative manner to make bis-thiazoles of the type found in myxothiazols and the related natural products **2** and **3**. More recently, Pd-catalysed cross-coupling reactions between 4-bromo-, 4-stannyl, and 4-triflyl thiazoles⁷ have been used to synthesise bis-thiazoles *en route* to cystothiazols.

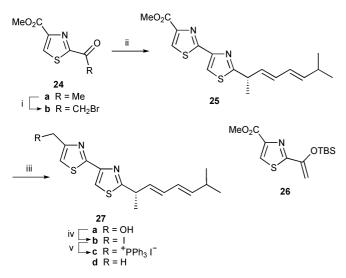
In our own studies we examined a variety of complementary routes to the bis-thiazole unit 7 in the myxothiazols.¹⁴ Ultimately, we found that a Hantzsch condensation between the thioamide **22** and the 2,4-disubstituted thiazole bromoketone **24b**, carried out under the conditions described by Holzapfel and Bredenkamp,¹⁵ provided the most reliable procedure for preparing the particular substituted bis-thiazole **25**.

The thiazole α -bromoketone **24b** is a known compound obtained by bromination of the corresponding methyl ketone **24a** using NBS in refluxing CCl₄.¹⁶ We experienced difficulty in



Scheme 1 Reagents and conditions: i) 'BuMe₂SiCl, imidazole (95%); ii) LiBH₄, THF (85%); iii) DMSO, $(COCl)_2$, $-78 \degree C$ (60%); iv) LiHMDS, **17**, $-78 \degree C$ (75%); v) Bu₄NF, THF (94%); vi) I₂, Et₂O, *hv* (74%); vii) Dess–Martin periodinane, NaHCO₃, then viii) NaClO₂, NaHPO₄ (61%); ix) (COCl)₂ then NH₃ (74%); x) Lawesson's reagent, THF (87%).

obtaining reproducible yields using these conditions, and ultimately produced the bromoketone **24b** by bromination of the intermediate enol silyl ether **26** derived from **24a**, using NBS in THF at 0 °C (Scheme 2). The bis-thiazole ester **25** was obtained as colourless prisms, with $[a]_D^{25} + 1.96$ (*c* 1.53 in CHCl₃). Reduction of the ester **25** using DIBAL-H in THF next gave the primary carbinol **27a**. Analysis of the corresponding Mosher's ester of **27a** gave an ee > 95%.



Scheme 2 Reagents and conditions: i) NBS, CCl_4 or NBS, THF then 26 (54%); ii) 24b and 22, NaHCO₃ at -20 °C, then TFA, pyridine (59%); iii) DIBAL-H, THF, -78 °C (56%); iv) I_2 , PPh₃, imidazole, 0 °C (78%); v) PPh₃, C_6H_6 (80%).

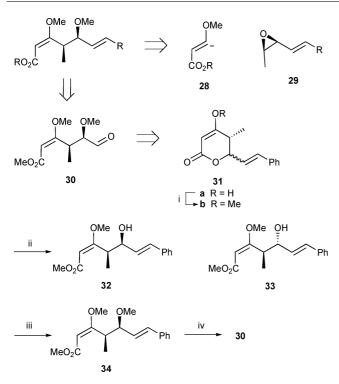
chain 6 to the bis-thiazole unit 7 using a Wittig reaction, we next treated the alcohol 27a with I_2/PPh_3 /imidazole at 0 °C under a nitrogen atmosphere, which led to the corresponding iodide 27b, as a solid, in 78% yield. Treatment of the iodide 27b with triphenylphosphine in benzene at room temperature then gave the corresponding phosphonium salt 27c, which was obtained as a colourless powder.

With the overall intention of linking the β -methoxyacrylate side

The substituted β -methoxyacrylate unit 6, and end game

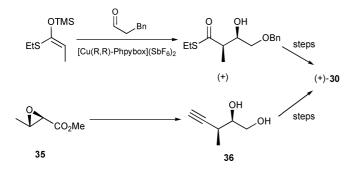
We discussed earlier the variety of synthetic approaches that have been applied, by others, to introduce the substituted β methoxyacrylate left-hand side chain, viz. 6, in endevours towards the related cystothiazols, *i.e.* 2 and 3. We examined a number of complementary synthetic routes to the β -methoxyacrylate side chain in myxothiazols in our studies, including those based on regioselective ring opening of chiral epoxides, *i.e.* 29, by both substituted lithium acetylides and vinyl anions e.g. 28, but to no avail. Ultimately, we decided to synthesise the aldehyde 30 and carry out a Wittig reaction with 27c leading to 1b. A synthetic approach to the β -methoxyacrylate aldehyde 30 that was attractive was via the 2H-pyran-2-one 31 (Scheme 3). The pyranone **31** was chosen since it incorporated a masked δ -hydroxy ester functionality, in addition to a β -methoxyacrylate residue with the required E-configuration. Furthermore, the styryl side chain in **31b** acted as precursor to the sensitive aldehyde functionality in the β -methoxyacrylate aldehyde **30**.

Thus, a condensation between cinnamaldehyde and the dianion produced from methyl 3-oxopentanoate,¹⁷ using NaH and *n*-BuLi in THF at 0 °C, first gave the pyran-2-one **31a** which, on methylation with $Me_2SO_4-K_2CO_3$, produced a 1 : 1 mixture of



Scheme 3 Reagents and conditions: i) Me_2SO_4 , K_2CO_3 (65%); ii) KOH, H_2O , 100 °C, then CH_2N_2 , then chromatography (9%); iii) MeI, Ag_2O (55%); iv) OsO_4 , NMO, Me_2CO-H_2O (9 : 1), then $NaIO_4$, $THF-H_2O$ (1 : 3) (65%).

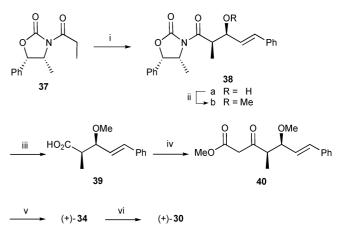
syn- and anti-diastereoisomers of the corresponding methyl ether 31b. The diastereoisomers of 31b could be separated by HPLC and their relative stereochemistries followed from inspection of the magnitude of their vicinal coupling between C5 and C6 in the ¹H NMR spectra, and comparison of these data with those obtained for similar compounds in the literature.¹⁸ Thus, the syndiasteroisomer of 31b displayed vicinal coupling of 3.6 Hz between the hydrogen atoms at C5 and C6 whereas the corresponding anti-diastereoisomer showed vicinal coupling of 7.1 Hz between the same hydrogens. For the related compound kawain-5-ol the corresponding vicinal couplings have been recorded as 3.0 Hz (syndiastereoisomer) and 7.0 Hz (anti-diastereoisomer).18 Preparative HPLC separation of the diastereoisomers of 31b was avoided in the synthetic scheme at this point, however, and instead the mixture of diastereoisomers was heated with 1.2 equivalents of KOH in H₂O at 100 °C followed by treatment with diazomethane leading to the syn and anti β -methoxyacrylate secondary alcohols 32 and 33 respectively, which could be easily separated by routine column chromatography. Methylation of the syn-diastereoisomer 32 using



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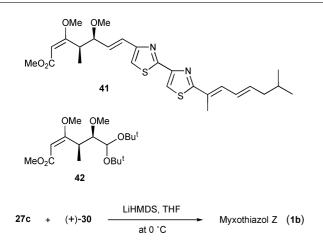
MeI and Ag₂O in diethyl ether finally led to the 3*RS*, 5*RS* methyl ether diastereoisomer **34** as a colourless oil. Oxidative cleavage of **34** *via* the corresponding isolated intermediate vicinal diol, using OsO_4 -NMO, followed by $NaIO_4$, finally gave the (±)- β -methoxyacrylate aldehyde **30**, as a labile liquid.

The same β -methoxyacrylate aldehyde 30 was later synthesised by Backhaus¹⁹ using aldol chemistry starting from benzyloxyacetaldehyde, and Deroy and Charette^{7b} developed an Evans enantiopure asymmetric aldol protocol to afford the (+)-aldehyde **30**. Akita *et al.*^{7*a*} also developed a route to enantiopure **30** from the chiral acetylenic alcohol 36 produced from ring opening of the 2R, 3S epoxy butanoate 35. We also synthesised the enantiopure β -methoxyacrylate aldehyde 30 by first using an Evans aldol reaction between the auxiliary 37 and cinnamaldehyde, which gave the syn-aldol 38a with >98% ee (Scheme 4).²⁰ O-Methylation of 38a, followed by conversion of the intermediate methyl ether 38b into the corresponding carboxylic acid 39 and treatment of the latter with LiCH₂CO₂Me next led to the β ketoester 40. Deprotonation of the β -ketoester 40, using NaH in DMPU, followed by addition of dimethyl sulfate then led to the enantiopure (+)- β -methoxyacrylate 34. Oxidative cleavage of the styryl alkene bond in 34, using the same conditions as those used with the racemic material, finally gave the $(+) \beta$ -methoxyacrylate aldehyde **30**, as an oil $[a]_{D}^{25} + 105$ (*c* 0.55 in CHCl₃).



Scheme 4 Reagents and conditions: i) n-Bu₂BOTf, 37, then PhCH=CH-CHO (90%); ii) 2,6-'Bu₂C₃H₃N, MeOTf (72%); iii) LiOH, H₂O₂ (98%); iv) NNCD, then LiCH₂CO₂Me, -78 °C (85%); v) NaH, Me₂SO₄, DMPU (66%); vi) OsO₄, NMO, Me₂CO–H₂O (9 : 1), then NaIO₄, THF–H₂O (1 : 3) (65%). (DMPU = *N*,*N*'-dimethylpropylene urea; NNCD = 2-chloro-4-nitro-1-benzenediazonium 2-naphthalenesulfonate.)

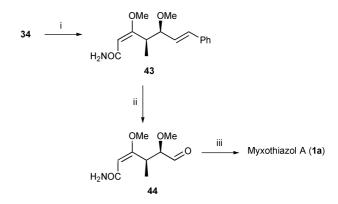
With both the phosphonium salt 27c, and racemic and enantiopure β -methoxyacrylate aldehyde 30 in hand, we were now in a position to examine their coupling to give myxothiazol Z (1b). This Wittig reaction between 27c and 30 proved to be somewhat temperamental. For example, when the salt 27c was treated with KOBu' in THF at 0 °C followed by addition of the (±)-aldehyde 30, only the product 27d resulting from hydrolysis of the phosphonium salt was isolated. Furthermore, when a mixture of 27c and (±)-30 was treated with KOBu' in THF only the positional isomer 41 of myxothiazol Z and the di-*t*-butyl acetal 42 of the aldehyde 30 were isolated. After more experimentation, use of the less basic



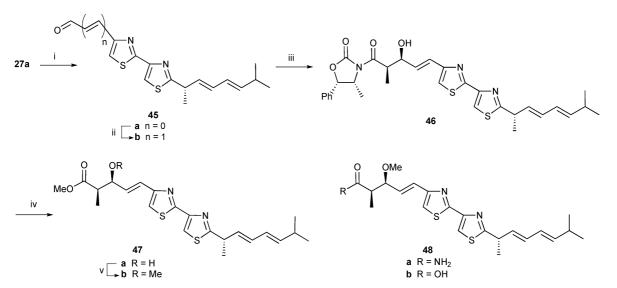
sodium methoxide in THF at 0 °C finally delivered myxothiazol Z (**1b**), which was obtained as a single alkene geometrical isomer after HPLC purification, in 11% yield. The synthetic myxothiazol showed spectroscopic data which were identical to those described for myxothiazol Z isolated from myxobacteria, even though this structure was not published as a natural product until some time after the completion of this work. A corresponding Wittig reaction between the bis-thiazole salt **27c** and the enantiopure (+)-aldehyde **30**, using lithium hexamethyldisilazide as the base in THF at 0 °C, proceeded much more smoothly and led to (+)-myxothiazol Z in 74% yield. A small amount of the corresponding Z-alkene was produced concurrently during the condensation which could be largely removed by HPLC. The synthetic myxothiazol Z had $[a]_{D}^{25}$ +118.8 (*c* 1.44 in CHC1₃) whereas Ahn *et al.* give $[a]_{D}^{25}$ +152, and Hőfle and co-workers $[a]_{D}^{22}$ +79.2 for naturally derived material.²

Attempts were now made to convert myxothiazol Z (**1b**) into myxothiazol A (**1a**) using Weinreb's conditions, *i.e.* Me₂AlNH₂ in CH₂Cl₂ However, these attempts failed, and instead only starting material was recovered. However, treatment of the β methoxyacrylate **34** with Me₂AlNH₂ gave the corresponding amide **43** in 40% yield (Scheme 5). Oxidative cleavage of the styryl alkene bond in **43**, using OsO₄–NaIO₄ then gave the β - methoxyacrylamide aldehyde 44. Gratifyingly, a Wittig reaction between this aldehyde and the phosphonium salt 27c using LiHMDS in THF at 0 °C was found to be *E*-selective and led to 7*R*, 18*SR*, 19*RS* myxothiazol A (1a), albeit in only 12% yield. The synthetic myxothiazol A showed spectroscopic data which were completely superimposable on those of the natural product derived from *M. fulvus*.

As a corollary to the aforementioned synthetic work, we also examined a synthetic approach to the bis-thiazole metabolites **48a** and **48b** known as myxothiazol R and myxothiazol S respectively, which co-occur with the β -methoxyacrylate myxothiazols A and Z in myxobacteria.²¹ Thus, a Wittig reaction between the bisthiazole aldehyde **45a** produced from the alcohol **27a**, and (formylmethylene) triphenylphosphorane first gave the corresponding α , β -unsaturated aldehyde **45b** as a pale yellow solid in 72% yield (Scheme 6). An Evans aldol condensation between **37** and the aldehyde **45b** in the presence of *n*-Bu₂BOTf next gave the β -hydroxy amide **46**, which was then converted into the methyl ester **47a** using MeMgBr in MeOH at 0 °C. Treatment of **47a** with NaOH–MeI gave the corresponding methyl ether **47b** which is the methyl ester analogue of the amide **48a**, *i.e.* myxothiazol R, and the carboxylic



Scheme 5 Reagents and conditions: i) Me_2AINH_2 , CH_2CI_2 (40%); ii) OsO_4 , Me_2CO-H_2O (9 : 1), then $NaIO_4$, $THF-H_2O$ (1 : 3) (32%); iii) LiHMDS, THF added to 44 and 27c (22%).



Scheme 6 Reagents and conditions: i) PDC, CH_2Cl_2 (58%); ii) OHC·CH=PPh₃, C_6H_6 (72%); iii) **37**, *n*-Bu₂BOTf, Et₃N, 0 °C, then **45** (93%); iv) MeMgBr, MeOH, 0 °C (52%); v) MeI, NaOH, DMSO-H₂O (2 : 1) (45%).

acid **48b**, *i.e.* myxothiazol S. Unfortunately, the dearth of synthetic materials did not allow us to realise the syntheses of myxothiazols R and S from the methyl ester **47b**.

Acknowledgements

We thank the SERC (now EPSRC) for studentships (CASE awards to B. J. M. and I. R. W.) and the Commonwealth Scholarship Commission for a scholarship (to H. D.). We also thank Dr G. Höfle (GBF, Braunshweig, Germany) for correspondence and exchange of information on natural myxothiazols.

References

- K. Gerth, H. Irschik, H. Reichenbach and W. Trowitzsch, J. Antibiot., 1980, 33, 1474–1479; W. Trowitzsch, G. Reifenstahl, V. Wray and K. Gerth, J. Antibiot., 1980, 33, 1480–1490; W. Trowitzsch, G. Höfle and W. S. Sheldrick, Tetrahedron Lett., 1981, 22, 3829–3832; W. Kohl, B. Witte, B. Kunze, V. Wray, D. Schomburg, H. Reichenbach and G. Höfle, Liebigs Ann. Chem., 1985, 2088–2097; W. Trowitzsch-Kienast, V. Wray, K. Gerth, H. Reichenbach and G. Höfle, Liebigs Ann. Chem., 1986, 93–98.
- 2 J.-W. Ahn, S.-H. Woo, C. O. Lee, K.-Y. Cho and B.-S. Kim, J. Nat. Prod., 1999, 62, 495–496; H. Steinmetz, E. Forche, H. Reichenbach and G. Höfle, *Tetrahedron*, 2000, 56, 1681–1684.
- 3 M. Ojika, Y. Suzuki, A. Tsukamoto, Y. Sakagami, R. Fudou, T. Yoshimura and S. S. Yamanaka, J. Antibiot., 1998, 51, 275–282; B. Böhlendorf, M. Herrmann, H. J. Hecht, F. Sasse, E. Forche, B. Kunze, H. Reichenbach and G. Höfle, Eur. J. Org. Chem., 1999, 2601–2608.
- 4 T. Anke, H.-J. Hecht, G. Schramm and W. Steglich, J. Antibiot., 1979, 32, 1112–1117; W. F. Becker, G. Von Jagow, T. Anke and W. Steglich, FEBS Lett., 1981, 132, 329–333; T. Anke, H. Besl, U. Mocek and W. Steglich, J. Antibiot., 1983, 36, 661–666; S. Backens, W. Steglich, J. Baeuerle and T. Anke, Liebigs Ann. Chem., 1988, 5, 405–409; T. Anke, G. Schramm, B. Schwalge, B. Steffan and W. Steglich, Liebigs Ann. Chem., 1984, 1616–1625.
- 5 J. M. Clough, *Nat. Prod. Rep.*, 1993, **10**, 565–574; G. Von Jagow and A. Link, *Methods Enzymol.*, 1986, **126**, 253–271; H. Augustiniak, K. Gerth, L. Grotjahn, H. Irschik, T. Kemmer, B. Kunze, H. Reichenbach, G. Reifenstahl, W. Trowitzsch and V. Wray, *Ger. Pat.*, 1978, DE 2838542 (*Chem. Abstr.*, 1980, **92**, 179082c).
- 6 D. W. Bartlett, J. M. Clough, J. R. Godwin, A. A. Hall, M. Hamer and B. Parr-Dobrzanski, *Pestic. Manage. Sci.*, 2002, 58, 649–662; D. W.

Bartlett, J. M. Clough, C. R. A. Godfrey, J. R. Godwin, A. A. Hall, S. P. Heaney and S. J. Maund, *Pestic. Outlook*, 2001, **12**, 143–148; H. Sauter, W. Steglich and T. Anke, *Angew. Chem., Int. Ed.*, 1999, **38**, 1328–1349.

- 7 (a) K. Kato, A. Nishimura, Y. Yamamoto and H. Akita, *Tetrahedron Lett.*, 2002, 43, 643–645; (b) P. L. Deroy and A. B. Charette, Org. Lett., 2003, 5, 4163–4165; (c) D. R. Williams, S. Patnaik and M. P. Clark, J. Org. Chem., 2001, 66, 8463–8469; (d) T. Bach and S. Heuser, Angew. Chem., Int. Ed., 2001, 40, 3184–3185; (e) J. Shao and J. S. Panek, Org. Lett., 2004, 6, 3083–3085; (f) M. Ojika, T. Watanabe, J. Qi, T. Tanino and Y. Sakagami, Tetrahedron, 2004, 60, 187–194; (g) P. Stanetty, M. Schnürch and M. D. Mihovilovic, J. Org. Chem., 2006, 71, 3754–3761.
- 8 B. J. Martin, J. M. Clough, G. Pattenden and I. R. Waldron, *Tetrahedron Lett.*, 1993, **34**, 5151–5154.
- 9 J. B. Baudin, G. Hareau, S. A. Julia and O. Ruel, *Tetrahedron Lett.*, 1991, 32, 1175–1178; J. B. Baudin, G. Hareau, S. A. Julia and O. Ruel, *Bull. Soc. Chim. Fr.*, 1993, 130, 336–357; J. B. Baudin, G. Hareau, S. A. Julia, R. Lorne and O. Ruel, *Bull. Soc. Chim. Fr.*, 1993, 130, 856–878; N. D. Smith, P. J. Kocienski and S. D. A. Street, *Synthesis*, 1996, 652–666.
- 10 M. P. Green, J. C. Prodger and C. J. Hayes, *Tetrahedron Lett.*, 2002, 43, 6609–6611.
- 11 J. A. Dale, D. L. Dull and H. S. Mosher, J. Org. Chem., 1969, 34, 2543–2549.
- 12 A. Hantsch and H. J. Weber, Berichte, 1887, 20, 3118-3124.
- 13 D. A. McGowan, U. Jordis, D. K. Minster and S. M. Hecht, J. Am. Chem. Soc., 1977, 99, 8078–8079.
- 14 I. R. Waldron, PhD Thesis, University of Nottingham, 1987; S. Comins, PhD Thesis, University of Nottingham, 1990; J. B. Martin; PhD Thesis, University of Nottingham, 1994.
- 15 M. W. Bredenkamp, C. W. Holzapfel and W. J. van Zyl, Synth. Commun., 1990, 20, 2235–2249.
- 16 T. T. Sakai, J. N. Riordan, T. E. Booth and J. D. Glickson, J. Med. Chem., 1981, 24, 279–285.
- 17 cf. H. Meyer and D. Seebach, *Liebigs Ann. Chem.*, 1975, 2261–2278;
 T. Reffstrup and P. M. Boll, *Acta Chem. Scand. Ser. B*, 1976, **30**, 613–618.
- 18 R. Hansel and J. Schultz, Chem. Ber., 1973, 106, 570-575.
- 19 D. Backhaus, Tetrahedron Lett., 2000, 41, 2087-2090.
- 20 D. A. Evans, J. Bartroli and T. L. Shih, J. Am. Chem. Soc., 1981, 103, 2127–2129; D. A. Evans, J. M. Takacs, L. R. McGee, M. D. Ennis, D. J. Mathre and J. Bartroli, *Pure Appl. Chem.*, 1981, 53, 1109–1127; J. R. Gage and D. A. Evans, *Org. Synth.*, 1990, 68, 83–91 and refs. cited therein.
- 21 N. Bedorf, B. Kunze, H. Reichenbach and G. Höfle, in *Scientific Annual Report of the Gesellschaft für Biotechnologische Forschung mbH*, Braunschweig, Germany, 1986, p. 14.